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Quantifying The Individual Impact Of Artificial Barriers In Freshwaters: A Standardized And Absolute Index Of Genetic Connectivity

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2	FRESHWATERS:
3	A STANDARDIZED AND ABSOLUTE INDEX OF GENETIC CONNECTIVITY
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22 Abstract

23 Fragmentation by artificial barriers is an important threat to freshwater biodiversity. Mitigating the 24 negative aftermaths of fragmentation is of crude importance, and it is now essential for environmental managers to benefit from a precise estimate of the individual impact of weirs and dams on river 25 26 connectivity. Although the indirect monitoring of connectivity using molecular data constitutes a promising approach, it is still plagued with several constraints preventing a standardized and 27 individual quantification of barrier effects. Indeed, observed levels of genetic differentiation depend 28 29 on both the age of the obstacle and the effective size of the populations it separates, making difficult 30 comparisons among obstacles. Here, we developed a standardized index of genetic connectivity 31 (C_{INDEX}) , allowing an absolute and independent assessment of the individual effects of obstacles on connectivity. The C_{INDEX} is the standardized ratio (expressed as a percentage) between the observed 32 33 genetic differentiation between pairs of populations located on either side of an obstacle and the 34 genetic differentiation expected if this obstacle completely prevented gene flow. The expected genetic 35 differentiation is calculated from simulations taking into account both the age of the barrier and the 36 effective size of the targeted populations. Using both simulated and published empirical datasets, we 37 explored and discussed the validity and the limits of the C_{INDEX} . We demonstrated that it allows quantifying genetic effects of fragmentation only a few generations after barrier creation and provides 38 39 valid comparisons among populations (or species) of different effective populations sizes and 40 obstacles of different ages. The computation of the C_{INDEX} requires a minimum amount of fieldwork and genotypic data, and solves some of the difficulties inherent to the study of artificial fragmentation 41 42 in rivers and potentially in other ecosystems. This makes the C_{INDEX} a promising and objective tool for managers aiming at restoring connectivity and at evaluating the efficiency of restoration programs. 43

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45 Keywords: Riverscape connectivity, Artificial fragmentation, Genetic differentiation, Simulations,

46 Bio-indicator, Dams, Weirs, Freshwater fish

47 Introduction

Heavily impacted by human activities, rivers are at the heart of biodiversity conservation issues 48 49 (Dudgeon et al., 2006; Reid et al., 2018). Among the various threats to these ecosystems, river fragmentation by artificial barriers is considered as the most widespread and worrying (Couto & 50 Olden, 2018; Nilsson, 2005; Turgeon, Turpin, & Gregory-Eaves, 2019). Weirs and dams, but also 51 52 pipes and culverts, have long been, and are still, constructed for flow regulation and/or hydropower 53 supply but they often imply a loss of habitat and a reduction in riverscape functional connectivity (that is, species-specific) in freshwater organisms (Birnie-Gauvin, Aarestrup, Riis, Jepsen, & Koed, 2017; 54 Jansson, Nilsson, & Malmqvist, 2007). For fish, artificial fragmentation is known to impact key 55 biological processes such as migration, dispersal and recruitment, and thus viability and productivity 56 57 of populations and communities (Blanchet, Rey, Etienne, Lek, & Loot, 2010; Poulet, 2007; Turgeon et al., 2019). Given the central role of hydropower as a source of energy, mitigating these negative 58 aftermaths is now of crude importance (Couto & Olden, 2018; Gibson, Wilman, & Laurance, 2017). 59

Different restoration and mitigation measures may be considered to enhance longitudinal river 60 61 connectivity, including the removal of obstacles, periodic turbine shutdowns and fishpasses setting 62 (Bednarek, 2001; Poff & Schmidt, 2016; Silva et al., 2018). However, these measures may all result in unintended outcomes (e.g., McLaughlin et al., 2013), or unsatisfactory trade-offs between 63 64 conservation of biodiversity, preservation of historical and cultural legacy and the maintenance of 65 services provided by obstacles (Gibson et al., 2017; Hand et al., 2018; Roy et al., 2018; Song et al., 66 2019). Benefiting from a precise estimate of the impact of an obstacle on river connectivity, or from a 67 precise estimate of the gain in connectivity resulting from a restoration action, is therefore essential for environmental managers and for conservation planning (Cooke & Hinch, 2013; Januchowski-Hartley, 68 69 Diebel, Doran, & McIntyre, 2014; Raeymaekers, Raeymaekers, Koizumi, Geldof, & Volckaert, 2009). 70 The direct monitoring methods conventionally used in rivers to quantify the functional permeability of 71 an obstacle or the efficiency of a restoration action are video-counting, telemetry and capture-72 recapture protocols. Although efficient (e.g., Cooke & Hinch, 2013; Hawkins et al., 2018; Junge et al., 73 2014; Pracheil et al., 2015), these methods are yet associated with technical constraints. In particular, ecological studies based on video counting or telemetry are often conducted on a limited number of
obstacles, whereas robust capture-recapture protocols imply repeated and exhaustive capture sessions,
ideally over several years, which involves the mobilization of substantial human and financial
resources (Cayuela et al., 2018).

78 Indirect monitoring based on molecular data constitutes a promising alternative approach, allowing 79 multi-specific studies of dam-induced fragmentation (Selkoe, Scribner, & Galindo, 2015). Among the 80 many analytical procedures developed in recent years to quantify the mobility of organisms on the 81 basis of genetic or genomic data, assignment methods and parentage analyses (Jombart, Devillard, & 82 Balloux, 2010; Pritchard, Stephens, & Donnelly, 2000; Städele & Vigilant, 2016; Wilson & Rannala, 83 2003) allow the detection of 'real-time' non-effective movements (that is, not necessarily followed by a reproduction event; e.g., Junge et al., 2014; Raeymaekers et al., 2009; Saint-Pé et al., 2018) but they 84 require an extensive sampling of individuals and moderate to high genetic differentiation between 85 populations (Broquet & Petit, 2009; Cayuela et al., 2018). 86

An alternative method to quantify the permeability of an obstacle from molecular data is simply to 87 88 measure the level of neutral genetic differentiation between populations located in the immediate upstream and downstream vicinity of an obstacle ("adjacent sampling strategy"), an approach that 89 does not necessarily require large sample sizes or heavy computation: any drop in local functional 90 91 connectivity due to the creation of a barrier to gene flow is expected to translate into an increase in 92 neutral genetic differentiation (Raeymaekers et al., 2009). However, measures of genetic differentiation may only be considered as correct estimates of barrier effects when comparing 93 94 obstacles of the same age and/or separating populations of similar effective size. This is because 95 genetic differentiation primarily stems from genetic drift, that is, from the random fluctuation of allelic 96 frequencies naturally occurring in all populations (Allendorf, 1986). When populations are separated 97 by an obstacle to gene flow, these fluctuations tend to occur independently in each population, leading 98 to a differential distribution of allelic frequencies on either side of the barrier. This process is yet 99 progressive, taking place over several generations (Landguth et al., 2010), and is all the more slow as effective population sizes are large (Broquet & Petit, 2009; Cayuela et al., 2018; Prunier, Dubut, 100 101 Chikhi, & Blanchet, 2017). As a consequence, it is impossible to attribute the differences in levels of 102 genetic differentiation observed between obstacles varying in age and/or in the effective size of 103 populations they separate to differences in their actual barrier effects; older obstacles or obstacles 104 separating smaller populations should show higher genetic differentiation than more recent obstacles 105 or obstacles separating larger populations, despite similar barrier effects. Given this drawback, there is 106 an urgent need for the development of a standardized and absolute index of genetic connectivity that 107 take into account the contribution of both the age of the obstacle and the effective size of populations 108 to observed measures of genetic differentiation. Such an index might allow a quick and robust 109 quantification of individual barrier effects whatever their characteristics, paving the way for informed 110 management prioritization and proper evaluation of restoration measures, along with inter-basins and 111 interspecific comparative studies.

Here, we bridge that gap by developing a user-friendly and standardized index of genetic connectivity, 112 allowing an absolute and independent assessment of the individual effects of obstacles on gene flow. 113 114 The proposed index (C_{INDEX}) is based on the relative comparison of observed measures of genetic differentiation resulting from an adjacent sampling strategy with theoretical machine learning 115 116 predictions obtained from genetic simulations. Genetic simulations are here used to reflect the expected evolution of allelic frequencies resulting from the interplay between the age of the obstacle 117 and the effective population sizes: the closer the observed measure of genetic differentiation from the 118 119 one that would be expected in the worst-case scenario (total barrier to gene flow), the lower the index 120 of connectivity. We first present the logic and principles underlying our index. We then use both 121 simulated and published empirical genetic datasets to explore and discuss the validity and the limits of 122 the proposed index. We finally propose several perspectives to use and further improve the index.

124 Material and methods

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126 Principle of the proposed index of genetic connectivity C_{INDEX}

127 The proposed index of genetic connectivity C_{INDEX} is designed as a standardized estimate of the 128 amount of gene flow that gets through an obstacle separating two adjacent populations. It simply 129 consists in rescaling the observed measure of genetic differentiation GD_{obs} within its theoretical range 130 of variation, taking into account the expected temporal evolution of allelic frequencies resulting from the interplay between the age of the obstacle and the averaged effective sizes of populations. This 131 theoretical range of variation spans from GD_{min} to GD_{max} . GD_{min} stands for the theoretical measure 132 of genetic differentiation that would be expected if the obstacle was totally permeable to gene flow 133 (migration rate $m \approx 0.5$). GD_{min} should theoretically equal 0 but the background noise resulting from 134 135 the concomitant influences of genetic drift, mutations and random sampling may actually lead to positive -yet very low- measures of genetic differentiation. On the other hand, GD_{max} stands for the 136 theoretical measure of genetic differentiation that would be expected under the worst-case scenario, 137 that is, under the hypothesis that the considered obstacle is a total barrier to gene flow (m = 0). GD_{max} 138 is expected to increase with time since barrier creation and to decrease with the increase in effective 139 140 population sizes (Gauffre, Estoup, Bretagnolle, & Cosson, 2008; Landguth et al., 2010). The index of 141 connectivity C_{INDEX} is then computed as follows (see Appendix S1 for details)

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$$C_{INDEX} = \left(\frac{\ln(GD_{obs}/GD_{max})}{\ln(GD_{min}/GD_{max})}\right) \times 100 = \left(\frac{\ln(GD_{obs}) - \ln(GD_{max})}{\ln(GD_{min}) - \ln(GD_{max})}\right) \times 100 \quad (\text{eqn. 1})$$

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145 It thus ranges from 0 % (the observed measure of genetic differentiation is maximum and equals the 146 expected value under the assumption that the considered obstacle acts as a total barrier to gene flow) to 147 100 % (the observed measure of genetic differentiation is minimum- but not null- and equals the 148 expected value under the assumption that the considered obstacle has no impact on gene flow). GD_{obs} 149 is directly calculated from observed genotypic data collected in populations located at the immediate 150 upstream and downstream vicinity of the obstacle, whereas GD_{min} and GD_{max} are predicted from theoretical datasets simulated according to three main parameters: the mutation rate μ of considered genetic markers, and (for GD_{max} only) the effective population size *Ne* of the two considered populations and the age T of the total barrier to gene flow (expressed in number of generations; e.g., Landguth et al., 2010; Lowe & Allendorf, 2010).

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156 Expected measures of genetic differentiation

We used QuantiNemo2 (Neuenschwander, Michaud, & Goudet, 2019), an individual-based simulator 157 for population genetics, to simulate theoretical datasets that will in turn be used to predict GD_{min} and 158 GD_{max} values. We designed a very simple meta-population composed of two adjacent demes, and we 159 used forward simulations of gene flow between these two demes over 1000 non-overlapping 160 generations. Each deme was initiated with Ne individuals and kept at a constant size over generations, 161 162 with Ne taking 93 different values ranging from 30 to 2000 individuals. Genetic polymorphism was 163 based on 15 microsatellite loci and 20 alleles per locus, which corresponds to the number of markers typically used in empirical study focusing on functional connectivity (e.g., Blanchet et al., 2010; 164 Coleman et al., 2018; Storfer, Murphy, Spear, Holderegger, & Waits, 2010). The mutation rate μ , 165 following a stepwise mutation model, was set to 5×10^{-5} or 5×10^{-4} , so as to explore the natural 166 variability observed in microsatellite markers (mutation rate ranging from 10⁻⁶ to 10⁻²; Li, Korol, 167 Fahima, Beiles, & Nevo, 2002; Schlötterer, 2000; Yue, David, & Orban, 2007). Genotypes were 168 randomly assigned to individuals at the beginning of simulations. The inter-deme migration rate was 169 set to 0.5 for the first 400 generations, a value providing an optimal mixing of populations (panmixia) 170 and mimicking a natural situation without barrier, and then dropped to zero for the last 600 171 generations, mimicking the creation of a total barrier to gene flow, splitting a "single" population into 172 two subpopulations. With populations being isolated for 600 generations, we made sure our 173 174 simulations covered a realistic time frame: most artificial barriers in freshwater ecosystems were 175 indeed built between the Middle Ages (12th–15th centuries) and today (Blanchet et al., 2010), which corresponds to a number of generations ranging from 0 to ~ 400 in aquatic organisms such as fish 176 species (assuming a generation time of 2 years for fish species). For each deme size Ne and each 177 178 mutation rate μ , we ran ten simulation replicates, and 30 genotypes were sampled every ten generations from generation 300 to generation 1000, resulting in a total of 132060 simulated genetic
datasets in the *Fstat* format (Goudet, 1995), further converted into the *genepop* format (Rousset, 2008)
using R (R Development Core Team, 2014).

For each dataset, we measured the two following pairwise metrics of genetic differentiation: the 182 Hedrick's G''st (Hedrick, 2005; Meirmans & Hedrick, 2011) and the Meirmans' of'st (Meirmans, 183 2006), both computed using the R-package mmod (Winter, 2012). Other metrics were initially 184 185 considered, but preliminary analyses indicated that some were dependent on sample size (e.g., the 186 proportion of shared alleles or the Cavalli-Sforza and Edwards' Chord distance; Bowcock et al., 1994; 187 Cavalli-Sforza & Edwards, 1967; see Appendix S2 for details), while others were sensitive to mutation 188 rate and/or did not show enough variability (e.g., the Weir and Cockerham's θ st or the Jost's D; Jost, 2008; Weir & Cockerham, 1984; see Appendix S3 for details): they were thus discarded to avoid 189 190 jeopardizing the validity of the proposed index. We found that the two retained metrics G''st and φ 'st 191 were robust to variations in mutation rate and increased quickly after barrier creation, especially in the 192 case of small effective population sizes (Appendix S3), in accordance with theoretical expectations 193 (Lowe & Allendorf, 2010; Meirmans & Hedrick, 2011). All negative G''st and φ 'st values were set to 0. In addition to these two measures of genetic differentiation G''st and φ 'st, we also computed the 194 195 averaged expected heterozygosity He over the 15 loci in each population. He was then averaged over 196 the two populations and further considered as a proxy for effective population sizes. Both theoretical 197 and empirical works indeed indicate that genetic diversity should increase with the increase in 198 effective population sizes (Hague & Routman, 2016; Kimura, 1983; see also Appendix S4). We here 199 focused on mean heterozygosity because, unlike metrics such as allelic richness, heterozygosity values 200 are bounded between 0 and 1, which facilitates comparison between case studies. Moreover, this 201 metric is much more straightforward to calculate for managers than the actual effective population 202 size, since the latter is notoriously difficult to estimate in complex landscapes (Paz-Vinas, Comte, et 203 al., 2013; Wang, 2005). These calculations resulted in a final dataset comprising 132060 lines and the 204 eight following columns: the simulated mutation rate μ , the generation t at which genotypes were recorded, the age T of the barrier computed as T = (t - 400) and expressed in number of generations 205 206 (the barrier being created at generation 400 in our simulations), the replicate number, the effective 207 population size *Ne* of each simulated population, the mean expected heterozygosity *He* and the two 208 metrics of genetic differentiation G''st and φ 'st.

209 A first subset of this final dataset was used as a training set in the regression implementation of a Random Forest machine-learning algorithm (Breiman, 2001). The objective was to establish 210 211 theoretical distributions allowing future predictions of GD_{max} values according to both time and effective population sizes (with *He* as a proxy). These theoretical distributions were designed so as to 212 mimic the temporal inertia in the setting up of genetic differentiation after the creation of a total barrier 213 214 to gene flow. For each mutation rate μ and each metric of genetic differentiation GD (either G''st or φ 'st) computed after the creation of the barrier (i.e., for T > 0), we used the R-package randomForest 215 (Liaw & Wiener, 2002) to fit the model $GD \sim T \times He$. We used 200 trees and a sample size of 500, as 216 these values provided very good accuracy (mean squared errors lower than 0.4%). Created 217 218 randomForest R-objects were saved in the form of .rda files (the usual file format for saving R 219 objects) and were further used to predict the four possible expected measures of genetic differentiation GD_{max} (two possible metrics of genetic differentiation and two possible mutation rates) between pairs 220 221 of populations according to both the mean expected heterozygosity He (the proxy for effective population sizes) and the number of generations T elapsed since barrier creation, using the 222 223 predict.randomForest function.

A second subset of the final dataset was used to predict the four possible measures of genetic differentiation GD_{min} (background signal) that may be expected under the influence of mutations, drift and random sampling between two adjacent populations not separated by any barrier to gene flow. For each of both mutation rates μ and each of both metrics of genetic differentiation GD (either G''st or φ 'st) computed before the creation of the barrier (i.e., for T < 0), GD_{min} was simply computed as the mean of simulated GD values. These four predicted GD_{min} values were stored in the form of a *.rda* file.

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232 Computing the index of connectivity C_{INDEX}

Equation 1 allows computing a unique index of connectivity C_{INDEX} for each combination of both a mutation rate μ (5×10⁻⁵ or 5×10⁻⁴) and a metric of genetic differentiation *GD* (G''st or φ 'st). The four indices are then averaged to get the final connectivity index C_{INDEX} with a 95% confidence interval computed as $1.96 \times SE$, with SE the estimated standard error (i.e., the estimated standard deviation divided by $\sqrt{4}$).

Note that when several genotypic datasets are available for a same obstacle, for instance when several 238 239 sympatric species are sampled on either side of the obstacle or when several replicates are considered 240 (as is the case of all simulated data in this study), an overall C_{INDEX} can also be estimated using an 241 intercept-only mixed-effect linear model with the various indices C as the response variable and the genotypic dataset as a random effect (Bates, Mächler, Bolker, & Walker, 2015). This procedure allows 242 243 taking into account the fact that indices of connectivity C_{INDEX} computed from the same dataset are not 244 independent and thus avoids biased estimates for standard error SE (McNeish, 2014). The overall 245 C_{INDEX} is then obtained from the estimated intercept of the model (which simply amounts to 246 calculating the average of indices C across datasets) and the corresponding 95% confidence interval is computed as $1.96 \times SE$, with SE the unbiased standard error as estimated by the mixed-effect model. 247

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The whole procedure was automated within a user-friendly R-function (see Appendix S10). Users are simply expected to provide an empirical genotypic dataset (in the *genepop* format) and a parameter file indicating for each considered obstacle the name of the two adjacent populations (as given in the genotypic dataset) and the number of generations elapsed since barrier creation. This number of generations is to be estimated from the life-history traits of the considered species. Figure 1 provides a flowchart allowing an overall visualization of the process.

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257 Validation of the connectivity index

To assess the validity of the proposed C_{INDEX} in response to different levels of obstacle permeability, we again used the program QuantiNemo2 to simulate gene flow between two demes over 1000 nonoverlapping generations. Demes were initiated with Ne = 50, 100, 250, 500 or 1000 individuals and kept at a constant size over generations. To mimic realistic genetic datasets, each microsatellite locus 262 was given a unique stepwise mutation rate μ randomly picked from a log-normal distribution ranging from 5×10^{-5} to 5×10^{-3} with a mean of 5×10^{-4} (see Appendix S5 for details). The inter-deme migration 263 264 rate was set to 0.5 for the first 400 generations, and then dropped to m for the last 600 generations, with *m* ranging from 0 to 0.2 with an increment of 0.01 and from 0.2 to 0.5 with an increment of 0.05, 265 mimicking the creation of a more or less severe barrier to gene flow (total barrier, m = 0; no barrier, m266 = 0.5). All other simulation parameters were similar to previous simulations. For each deme size Ne 267 268 and each migration rate m, we ran 20 simulation replicates, and 30 genotypes were sampled at generations 405 (age of the barrier T = 5), 410, 415, 420, 425, 450, 500 and 700 (T = 300), resulting in 269 270 a total of 21600 simulated genetic datasets in the *Fstat* format, further converted into the genepop 271 format.

For each simulated dataset, we computed the averaged expected heterozygosity He and the two 272 pairwise measures of genetic differentiation G''st and φ 'st. We then used parameters T and He to 273 predict the corresponding measures of genetic differentiation GD_{min} and GD_{max} (for both G''st and 274 φ 'st) expected under various mutation rates (5×10⁻⁵ and 5×10⁻⁴) using the *predict.randomForest* 275 function (Appendix S10) and the previously created .rda files. For each dataset, the four indices of 276 277 connectivity C were then computed using equation 1. To average datasets across replicates, we finally used intercept-only mixed-effect models (with dataset as a random effect) to get the final mean C_{INDEX} 278 (along with a 95% confidence interval) corresponding to each combination of N, T and m. 279

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281 Empirical data

To assess the behavior of the proposed index of connectivity in real situations, we used two published 282 empirical datasets. The first one is from Gouskov et al. (2016). In this study, authors assessed 283 riverscape fragmentation induced by 37 hydroelectric power stations in the Rhine catchment in 284 Switzerland using data from 2133 European chubs (Squalius cephalus) sampled across 47 sites and 285 genotyped at nine microsatellite loci. We selected 23 pairs of populations according to the following 286 criteria: upstream and downstream populations belonged to the same river, were separated by a single 287 dam, were distant from a maximum of 16km (maximum migration distance observed in chub 288 according to Fredrich et al., 2003) and were not separated by any confluence with tributaries. This 289

selection corresponded to 23 independent dams created between 1893 and 1970, all equipped with fishpasses (Table1; see also Appendix S6 for a map). We considered a generation time of 3 years, as reported in Gouskov et al. (2016) to compute the number of generations elapsed since barrier creation and ran our developed R-function to automatically compute C_{INDEX} values.

294 The second empirical dataset is from Prunier et al. (2018). In this study, authors assessed the influence 295 of various anthropogenic stressors including riverscape fragmentation induced by weirs on patterns of 296 genetic diversity and differentiation in two freshwater fishes from two distinct rivers in southwestern 297 France. They used data from 1361 Eurasian minnows (Phoxinus phoxinus) and 1359 Languedoc 298 gudgeon (Gobio occitaniae) sampled across 47 sites (22 in the Célé River and 25 in the Viaur River) 299 and genotyped at 11 and 13 microsatellite loci, respectively. We selected 8 pairs of populations 300 according to the following criteria: upstream and downstream populations belonged to the same river, 301 were separated by a single weir, were distant from a maximum of 1km, were not separated by any 302 confluence with tributaries and were sampled for both species. This selection corresponded to 8 303 independent weirs (~1 to 3 meters high) created between the 16th and the 20th century (Table 1; see 304 also Appendix S6 for maps). We considered a generation time of 2 years in P. phoxinus and 2.5 years 305 in G. occitaniae to compute the number of generations elapsed since barrier creation and again ran developed R-function (Appendix S10) to automatically compute C_{INDEX} values for each obstacle, each 306 307 species and across species.

309 **Results**

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311 Expected measures of genetic differentiation

The first set of simulations was designed to predict GD_{min} and GD_{max} values, that is, the lower and 312 upper limits of the theoretical range of variation of GD_{obs} . Data simulated before the creation of the 313 barrier (m = 0.5; T < 400) were used to predict GD_{min} values whereas data simulated after the creation 314 315 of the barrier (m = 0; T > 400) were used to predict GD_{max} values. As expected with a migration rate allowing panmixia, GD_{min} values were always very close from 0, ranging from 5.53×10^{-3} to 7.17×10^{-3} 316 ³ for G''st and from 7.46×10^{-3} to 8.51×10^{-3} for φ 'st. These values represent the predicted background 317 levels of genetic differentiation, resulting from the sole influences of random processes such as genetic 318 319 drift, mutations and sampling biases (Figure 2).

320 GD_{max} values were on the contrary designed to mimic the temporal inertia in the setting up of genetic 321 differentiation after the creation of a total barrier to gene flow. They were predicted from time since barrier creation and averaged expected heterozygosity (a proxy for effective population size) using a 322 Random Forest algorithm, simulated data being used as training sets. With explained variance ranging 323 324 from 86.8 to 94.2 %, Random Forest models accurately captured variations in measures of genetic 325 differentiation across the parameter space, whatever the considered mutation rate or the considered 326 metric of genetic differentiation (Appendix S7). As expected in absence of gene flow (Figure 2), GD_{max} increased with time since barrier creation and decreased with effective population size (*He*). 327 With predicted GD_{max} values ranging from 0.031 to 0.898 for G''st and from 0.042 to 0.968 for φ 'st, 328 both metrics displayed similar distribution patterns across mutation rates, although φ 'st systematically 329 330 showed higher values at low He.

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332 Validation of the connectivity index

The second set of simulations was designed to assess whether the C_{INDEX} correctly reflected the actual level of gene flow between two populations separated by an artificial barrier, beyond the temporal inertia in the setting-up of genetic differentiation. The mean C_{INDEX} values computed over simulated

replicates for each combination of Ne (effective population size), T (number generations since barrier 336 creation) and m (migration rate) showed –as expected- an overall increase with the increase in 337 338 migration rate, whatever the effective size of populations or the age of the barrier (Figure 3A-H; see Appendix S8 for a visualization against the number Nm of effective dispersal events per generation). 339 340 As expected when population are connected with high migration rates after the barrier setting (m > 0.2, a migration rate of 0.5 ensuring panmixia), the 95% confidence intervals for the C_{INDEX} always 341 342 included values higher than 90% (Figures 3A-H). On the contrary, in absence of gene flow after the 343 barrier setting (m = 0), the 95% confidence intervals always included values lower than 10%, except 344 within the first 5 generations after barrier creation (Figure 3A), or within the first 10 generations for very large effective population sizes (Figure 3B, $Ne \ge 1000$). In these cases, the C_{INDEX} was slightly 345 biased upwards, which indicates that we could not totally rule out the noise associated with the 346 347 measurement of genetic differentiation within the 5-10 first generations after barrier creation (Figure 348 3I; Appendix S9). Nevertheless, the C_{INDEX} showed valid and consistent values for both lowest and highest migration rates, the two thresholds of 10% (total barrier to gene flow) and 90% (full gene 349 350 flow) providing robust benchmarks for future interpretation of the index, whatever the age of the 351 obstacle (from generation 15 at least) or the effective size of populations.

For low migration rates ($0 < m \le 0.05$), the C_{INDEX} showed higher variability, with two noticeable 352 353 trends. First, all effective population sizes being combined, the C_{INDEX} slightly increased with the 354 increase in time since barrier creation (from generation 15 to generation 300), with a 10 to 20% 355 increase for lowest migration rates (but not for m = 0; Figure 3I). Secondly, all generations > 10 being 356 combined, the C_{INDEX} slightly increased with the increase in effective population sizes, with again a 10 357 to 20% increase for lowest migration rates from Ne = 50 to Ne = 1000 (Figure 3J). For instance, for m 358 = 0.05 after the barrier setting and for the most extreme cases, C_{INDEX} values ranged from 31.1 in 359 smallest populations to 50.1 in largest populations (mean = 41.2) at generation 15 (Figure 3C), and from 55.7 in smallest populations to 86.0 in largest populations (mean = 70.8) at generation 300 360 361 (Figure 3H).

362

363 Empirical data

In the first empirical dataset (Gouskov et al., 2016), monitored dams were created from 1893 to 1970, 364 which corresponds to approximately 13 to 39 generations in S. cephalus (Table1). Averaged levels of 365 366 expected heterozygosity were high and showed little variability (ranging from 0.69 to 0.77), whereas observed measures of genetic differentiation were pretty low, ranging from 0 to 0.032 for φ 'st and 367 from 0 to 0.018 for G''st. We found that six dams showed a C_{INDEX} value ranging from 64% to 75%, 368 suggesting from 25 to 36% decrease in genetic connectivity since barrier creation (Figure 4A). The 369 370 other 17 dams all showed C_{INDEX} values higher or equal to 90% (as indicated by confidence intervals), 371 indicating that populations located on either side of the barrier are properly connected by gene flow. 372 Importantly, C_{INDEX} values were independent from both time since barrier creation (spearman correlation test, $\rho = -0.18$, p = 0.42) and averaged heterozygosity ($\rho = 0.22$, p = 0.32) whereas both 373 φ 'st and G''st values tended to covary negatively with averaged heterozygosity (p < 0.09) and G''st 374 values to covary positively with time since barrier creation (p < 0.09). 375

376 In the second empirical dataset (Prunier et al., 2018), monitored weirs were built between the 16th and the 20th century, that is approximately from 20 to 204 generations in G. occitaniae and from 25 to 255 377 378 generations in P. phoxinus. As previously, averaged levels of expected heterozygosity were high and showed little variability (ranging from 0.58 to 0.72), whereas observed measures of genetic 379 differentiation were pretty low, ranging from 0 to 0.034 for φ 'st and from 0 to 0.026 for G''st. The 380 381 impact of weirs was variable across space and species (Table 1; Figure 4B). In P. phoxinus, all weirs 382 were found as highly permeable ($C_{INDEX} > 90\%$) with 7 out of 8 weirs showing a C_{INDEX} of 100%. In G. 383 occitaniae, four weirs were found as responsible for a decrease in genetic connectivity since barrier 384 creation ($C_{INDEX} < 90\%$), with C_{INDEX} values ranging from 83.9 (± 1.3%) in the case of barrier CAP in 385 the Viaur River to 49.2 (± 3.8 %) in the case of barrier SCC in the Célé River. When computed across 386 species, none of the weirs were identified as obstacles to overall genetic connectivity (C_{INDEX} ranging 387 from 77.2 to 100 %, with 95% confidence intervals systematically including the 90% threshold; Table 1). As previously, C_{INDEX} values were independent from both time from barrier creation ($\rho = 0.21$, p = 388 0.41) and averaged heterozygosity ($\rho = 0.18$, p = 0.47), but so were φ 'st and G''st values (p > 0.29). 389

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392 **Discussion**

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Restoring riverscape functional connectivity is of crude importance in terms of biodiversity 394 conservation and it is now often subject to regulatory obligations (e.g. in Europe, the Water 395 396 Framework Directive 2000/60/EC). However, rivers are subject to many and sometimes contradictory 397 uses (Reid et al., 2018): for practitioners to be able to propose informed trade-offs between restoring 398 riverscape connectivity and maintaining infrastructures and their associated socioeconomic benefits 399 (Hand et al., 2018; Roy et al., 2018; Song et al., 2019), new tools have to be developed, allowing a 400 rapid and reliable quantification of the relative impacts of obstacles to freshwater species movements. 401 Our objective was here develop an operational tool allowing such thorough quantification from a 402 minimum amount of data (Figure 1; see Box 1 for user guidelines).

The proposed index of genetic connectivity C_{INDEX} can be easily and automatically computed from a 403 404 simple set of upstream and downstream genotypes collected once and in the direct vicinity of a 405 putative barrier, provided the number of generations elapsed since barrier creation is known. Based on two complementary metrics of genetic differentiation (G''st and φ 'st) preliminary chosen so as to limit 406 any possible bias, the C_{INDEX} simply scales the observed level of genetic differentiation (GD_{obs}) with 407 408 respect to a theoretical range of variation spanning from the background noise expected in the absence of any barrier to gene flow ($GD_{min} \sim 0$, $C_{INDEX} = 100\%$) to the maximal level of differentiation 409 410 expected if the obstacle was a total barrier to gene flow (GD_{max} , $C_{INDEX} = 0\%$), the latter taking into 411 account both the time since barrier creation and the effective population size. Using numerous 412 simulations to explore the interplay between time since barrier creation, mutation rate and averaged expected heterozygosity (a proxy for effective population size), we were able to obtain GD_{max} values 413 for a large range of biologically realistic parameters (Figure 2). As expected, GD_{max} values 414 415 progressively increased with time since barrier creation and decreased with averaged expected 416 heterozygosity. Mutation rate also influenced GD_{max} patterns: as expected, higher mutation rates 417 accelerate genetic differentiation through time when population sizes are small to medium.

The C_{INDEX} showed constant patterns of increase with the increase in migration rates (from m = 0 to m418 = 0.2), whatever the number of generations since barrier creation and the effective population size 419 420 (Figure 3). For lowest migration rates (m ≤ 0.05), we yet found that it could underestimate barrier 421 effects in the first 5 to 10 generations after the creation of the obstacle. As a conservative strategy, we 422 suggest that the C_{INDEX} should not be used to assess the permeability of obstacles separating 423 populations for less than 10 generations. It is yet noteworthy that the C_{INDEX} can be applied to any type 424 of organisms and thus that species with low generation time (such as some invertebrate species) may 425 be considered as good candidates to investigate the impact of recently built barriers. For lowest 426 migration rates (m \leq 0.05), we also found that C_{INDEX} values slightly increased with both time since 427 barrier creation (from generations 15 to 300) and effective population sizes (from Ne = 50 to Ne=1000; Figure 3I-J). These trends have to be kept in mind when comparing intermediate C_{INDEX} values, 428 429 ranging from ~15 to ~70% (see Box 1 for an illustration).

430

431 Nevertheless, the C_{INDEX} provides a promising individual quantification of both the short- and long-432 term genetic effects of dam-induced fragmentation, allowing robust comparisons among species or 433 populations with different population sizes, and obstacles of different ages (from generation 15 at least) and types. When applied to empirical data, the C_{INDEX} allowed identifying several obstacles 434 435 partially limiting gene flow in at least two freshwater fish species (Figure 4): six out of the 23 dams monitored by Gouskov et al. (2016) and four out of the eight weirs monitored by Prunier et al. (2018) 436 437 showed a C_{INDEX} lower than 90%, in chubs and gudgeons, respectively. In each dataset, computed 438 C_{INDEX} values were systematically independent from both time from barrier creation and averaged heterozygosity, indicating that the C_{INDEX} properly takes into account the differential evolution of 439 440 allelic frequencies on either side of the barrier. Interestingly, the two most recent weirs in Prunier et al. 441 (2018; the SCC weir on the Célé River and the CIR weir on the Viaur River, both built in the 1960's, i.e. ~20 gudgeon generations ago; Table 1) showed contrasted results in gudgeons: the SCC weir was 442 443 identified as the most impactful obstacle ($C_{INDEX} = 49.2 \pm 3.8\%$), whereas the CIR weir was found as 444 highly permeable to gene flow ($C_{INDEX} = 86.7 \pm 15.1\%$). These contrasted results suggest that distinct typological features (height, slope, presence of a secondary channel, etc.) may differently affect fish 445

446 mobility (Baudoin et al., 2014). Furthermore, none of the monitored weirs were identified as barriers to gene flow in minnows, in accordance with personal field observations and previous findings on the 447 448 same two rivers (Blanchet et al., 2010). Although understanding how obstacle typological features and 449 fish traits might interact and shape riverscape patterns of functional connectivity was beyond the scope of this study, these results suggest that future comparative studies based on the proposed C_{INDEX} might 450 provide thorough insights as to the determinants of dam-induced fragmentation in various freshwater 451 452 organisms (Richardson, Brady, Wang, & Spear, 2016), including fish but also other taxa such as 453 macro-invertebrates that display very contrasting traits related to dispersal (e.g., Alp, Keller, Westram, & Robinson, 2012). 454

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456 Despite its strong operational potential, the C_{INDEX} does not come without some limitations. First of all, 457 it is important to remember that this index is a measure of genetic connectivity, not demographic 458 connectivity (Lowe & Allendorf, 2010), and thus cannot directly provide any counting of the actual number of crossing events. If immigrants do not reproduce, the actual crossing of dozens of 459 460 individuals, although suggesting high permeability, might not translate into high migration rates, resulting in low C_{INDEX} values (Figure 3). Furthermore, a migration rate has to be interpreted in regard 461 of effective population size: a migration rate of 0.05 actually corresponds to 2.5 effective dispersal 462 463 events per generation in populations of size 50, but to 50 effective dispersal events in populations of 464 size 1000 (Appendix S8). Despite higher permeability in the latter case, the contribution of these 50 465 immigrants to local recruitment might still be limited when compared to the contribution of residents 466 (Lowe & Allendorf, 2010). Although this is more of an inherent characteristic of the index than a real 467 limitation, it is important to keep this specificity in mind when interpreting it.

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469 Secondly, the computation of the C_{INDEX} relies on the assumption that, beyond the background signal 470 of genetic differentiation that is expected under the sole influences of genetic drift, mutations and 471 random sampling (GD_{min}) , the observed measures of genetic differentiation GD_{obs} only stem from 472 dam-induced fragmentation. This assumption is only valid when sampling adjacent populations, 473 located at the immediate upstream and downstream vicinity of the considered obstacle (Figure 1). It 474 thus implies the exclusion of migratory fish species, though at the heart of great conservation issues (e.g., Junge et al., 2014; Klütsch et al., 2019): complex life-cycles such as anadromy ("river-sea-river" 475 476 migrations), catadromy ("sea-river-sea" migrations) or potamodromy ("river-lake-river" migrations) 477 indeed preclude the delineation of upstream and downstream populations and do not allow proper 478 estimates for the C_{INDEX} . In non-migratory fish species, this assumption also limits the use of the C_{INDEX} 479 in large-scale studies, in which the distance between the upstream and the downstream sampling sites 480 lies beyond the dispersal capacities of the studied species. It certainly leaves room for manoeuvre, as 481 illustrated with the empirical dataset from Gouskov et al. (2016): we could for instance select pairs of 482 populations located up to 16km apart, but this was only possible because of the high mobility of 483 chubs, and performed in an illustrative purpose. In low-mobility species for instance, a non-adjacent 484 sampling might bias the C_{INDEX} downwards and hence overestimate the measure of fragmentation, as 485 observed measures of genetic differentiation would result from dam-induced fragmentation but also 486 from other processes such as Isolation-by-Distance (e.g. Coleman et al., 2018). We thus strongly 487 encourage practitioners to consider an adjacent sampling design as often as possible, although we 488 readily acknowledge that this may not always be an easy task given safety and accessibility considerations. Furthermore, fish may not always be found in the direct vicinity of obstacles. For 489 instance, the conversion of a river into a reservoir after the creation of a dam often leads to major 490 491 habitat modification and shifts in species composition (Bednarek, 2001), which can force adapting the 492 sampling design. A solution might be to capture the resultant background signal of genetic 493 differentiation by simulating GD_{min} values under various scenarios of isolation (Isolation-by-Distance, 494 Isolation-by-Resistance, etc.; McRae, 2006) in a way similar to the simulation of GD_{max} values in this 495 study (Figure 2). We yet believe that the variety, the complexity and the specificity of such scenarios 496 would preclude the computation of standardized C_{INDEX} scores, comparable among obstacles, species and studies. Although it might in some instances be considered a technical constraint, we argue that 497 498 only a strict adjacent sampling design can warrant unbiased and reliable C_{INDEX} estimates.

499

Finally, the proposed C_{INDEX} does not take into account the possible asymmetric gene flow created by barriers, as fish might struggle or even fail to ascent an obstacle (sometimes despite the presence of 502 dedicated fishpasses; Silva et al., 2018) whereas dam discharge might on the contrary further increase 503 or even force downstream movements (Pracheil et al., 2015). Although quantifying the asymmetric 504 permeability of obstacles appears of crude importance for informed conservation measures, the 505 proposed C_{INDEX} currently relies on the use of classical pairwise measures of genetic differentiation 506 that assume symmetric gene flow. Future developments will be required to allow the C_{INDEX} to provide 507 unbiased and distinct standardized scores for both upstream and downstream barrier effects. In the 508 meanwhile, it may be interesting to also assess the validity of the C_{INDEX} in quantifying the effects of 509 terrestrial obstacles, since asymmetric gene flow is not necessarily as pronounced as in river systems: provided that populations are sampled in the direct vicinity of the obstacle, the C_{INDEX} might as well 510 provide a standardized quantification of road-induced fragmentation. 511

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514 Conclusion

We here laid the groundwork for an operational tool dedicated to the individual and standardized 515 516 quantification of the impact of artificial barriers on riverscape functional connectivity from measures 517 of genetic differentiation. The proposed index of genetic connectivity C_{INDEX} is designed to take into account the temporal inertia in the evolution of allelic frequencies resulting from the interplay between 518 519 the age of the obstacle and the effective sizes of populations. Provided only adjacent populations are 520 sampled, the C_{INDEX} allows a rapid and thorough ranking of obstacles only a few generations after their 521 creation. The C_{INDEX} in its current form still suffers from some limitations, and it should be seen as the preliminary version of a future powerful bio-indicator of habitat fragmentation, rather than as an end-522 product. We call conservation and population geneticists to pursue the development of such an index, 523 524 as we -as scientists- need to help managers resolve complex and urging social problems. Nonetheless, 525 the C_{INDEX} is robust, only requires a minimum amount of fieldwork and genotypic data and already 526 solves several difficulties inherent to the study of dam-induced fragmentation in river systems, making 527 it a promising tool for the restoration of riverscape connectivity. The C_{INDEX} may allow practitioners to 528 objectively identify obstacles that do not present any substantial conservation issue (from a connectivity perspective) and help them target their efforts and resources towards the most impactful ones. Similarly, it may allow tracking the expected temporal decrease in genetic differentiation after obstacle removal or fishpass setting (Landguth et al., 2010; Schwartz, Luikart, & Waples, 2007) and thus help evaluate the success of local mitigations and restoration measures in response to regulatory obligations (Silva et al., 2018). Finally, it might as well provide a standardized quantification of roadinduced fragmentation, a critical issue in terrestrial ecology.

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537 Acknowledgements

We warmly thank all the colleagues and students who helped with field sampling. We are also grateful to Dr. A. Gouskov and C. Vorburger for details about their data. This work has been financially supported by grants to SB from the Agence Française pour la Biodiversité and from the Région Occitanie (CONAQUAT).

542

543 Supporting Information

544

545 Appendix S1: Details on the C_{INDEX} formula.

546 Appendix S2: Effect of sample size on various measures of genetic differentiation.

547 Appendix S3: Effect of both mutation rate and time since barrier creation on various measures of

548 genetic differentiation.

549 Appendix S4: Relationship between mean expected heterozygosity and mean effective population size.

550 Appendix S5: Distribution of mutation rates used in the second set of simulations.

551 Appendix S6: Localization of obstacles in the two empirical datasets.

552 Appendix S7: Relationship between observed and predicted measures of genetic differentiation.

553 Appendix S8: C_{INDEX} responses to the increase in the number of effective dispersal events (*Nm*).

- 554 Appendix S9: Rational for the observed bias in C_{INDEX} values in the very first generations after barrier
- 555 creation.
- 556 Appendix S10: Walkthrough for the computation of the C_{INDEX}
- 557 Appendix S11: https://doi.org/10.6084/m9.figshare.9698879.v1 (R-objects for the computation of the
- 558 C_{INDEX})

559 **REFERENCES**

- Allendorf, F. W. (1986). Genetic drift and the loss of alleles versus heterozygosity. *Zoo Biology*, *5*(2),
 181-190. (WOS:A1986C753400010).
- 562 Alp, M., Keller, I., Westram, A. M., & Robinson, C. T. (2012). How river structure and biological traits
- 563 influence gene flow : A population genetic study of two stream invertebrates with differing
- 564 dispersal abilities: Biological traits and gene flow in stream invertebrates. *Freshwater Biology*,

565 57(5), 969-981. doi: 10.1111/j.1365-2427.2012.02758.x

- Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting Linear Mixed-Effects Models Using
 Ime4. Journal of Statistical Software, 67(1). doi: 10.18637/jss.v067.i01
- 568 Baudoin, J.-M., Burgun, V., Chanseau, M., Larinier, M., Ovidio, M., Sremski, W., ... Voegtle, B. (2014).
- 569 The ICE protocol for ecological continuity. Assessing the passage of obstacles by fish.

570 *Concepts, design and application*. Paris: Onema.

571 Bednarek, A. T. (2001). Undamming Rivers : A Review of the Ecological Impacts of Dam Removal.

572 Environmental Management, 27(6), 803-814. doi: 10.1007/s002670010189

- 573 Birnie-Gauvin, K., Aarestrup, K., Riis, T. M. O., Jepsen, N., & Koed, A. (2017). Shining a light on the loss
- of rheophilic fish habitat in lowland rivers as a forgotten consequence of barriers, and its
- 575 implications for management. Aquatic Conservation: Marine and Freshwater Ecosystems,
- 576 27(6), 1345-1349. doi: 10.1002/aqc.2795
- 577 Blanchet, S., Rey, O., Etienne, R., Lek, S., & Loot, G. (2010). Species-specific responses to landscape 578 fragmentation : Implications for management strategies. *Evolutionary Applications*, *3*(3),
- 579 291-304. doi: 10.1111/j.1752-4571.2009.00110.x
- 580 Bowcock, A. M., Ruizlinares, A., Tomfohrde, J., Minch, E., Kidd, J. R., & Cavallisforza, L. L. (1994). High-
- resolution of human evolutionary trees with polymorphic microsatellites. *Nature*, *368*(6470),
- 582 455-457. (ISI:A1994ND12000063).
- 583 Breiman, L. (2001). Random forests. *Machine Learning*, 45(1), 5-32. (WOS:000170489900001).

- 584 Broquet, T., & Petit, E. J. (2009). Molecular Estimation of Dispersal for Ecology and Population
- 585 Genetics. *Annual Review of Ecology, Evolution, and Systematics, 40*(1), 193-216. doi:

586 10.1146/annurev.ecolsys.110308.120324

- 587 Cavalli-Sforza, L. L., & Edwards, A. W. (1967). Phylogenetic analysis. Models and estimation
 588 procedures. *American journal of human genetics*, *19*, 233.
- 589 Cayuela, H., Rougemont, Q., Prunier, J. G., Moore, J.-S., Clobert, J., Besnard, A., & Bernatchez, L.
- 590 (2018). Demographic and genetic approaches to study dispersal in wild animal populations :
- 591 A methodological review. *Molecular Ecology*, 27(20), 3976-4010. doi: 10.1111/mec.14848
- 592 Coleman, R. A., Gauffre, B., Pavlova, A., Beheregaray, L. B., Kearns, J., Lyon, J., ... Sunnucks, P. (2018).
- 593 Artificial barriers prevent genetic recovery of small isolated populations of a low-mobility
- 594 freshwater fish. *Heredity*. doi: 10.1038/s41437-017-0008-3
- 595 Cooke, S. J., & Hinch, S. G. (2013). Improving the reliability of fishway attraction and passage
- efficiency estimates to inform fishway engineering, science, and practice. *Ecological Engineering*, *58*, 123-132. doi: 10.1016/j.ecoleng.2013.06.005
- Couto, T. B., & Olden, J. D. (2018). Global proliferation of small hydropower plants—Science and
 policy. *Frontiers in Ecology and the Environment*. doi: 10.1002/fee.1746
- 600 Dudgeon, D., Arthington, A. H., Gessner, M. O., Kawabata, Z.-I., Knowler, D. J., Lévêque, C., ... Sullivan,
- 601 C. A. (2006). Freshwater biodiversity : Importance, threats, status and conservation

602 challenges. *Biological Reviews*, *8*1(2), 163-182. doi: 10.1017/S1464793105006950

- Fredrich, F., Ohmann, S., Curio, B., & Kirschbaum, F. (2003). Spawning migrations of the chub in the
- 604 River Spree, Germany. Journal of Fish Biology, 63(3), 710-723. doi: 10.1046/j.1095-
- 605 8649.2003.00184.x
- Gauffre, B., Estoup, A., Bretagnolle, V., & Cosson, J. F. (2008). Spatial genetic structure of a small
- 607 rodent in a heterogeneous landscape. *Molecular Ecology*, 17(21), 4619-4629. doi:
- 608 10.1111/j.1365-294X.2008.03950.x

- Gibson, L., Wilman, E. N., & Laurance, W. F. (2017). How Green is 'Green' Energy? *Trends in Ecology & Evolution*, *32*(12), 922-935. doi: 10.1016/j.tree.2017.09.007
- Goudet, J. (1995). FSTAT (Version 1.2) : A Computer Program to Calculate F-Statistics. *Journal of Heredity*, *86*(6), 485-486. doi: 10.1093/oxfordjournals.jhered.a111627
- 613 Gouskov, A., Reyes, M., Wirthner-Bitterlin, L., & Vorburger, C. (2016). Fish population genetic
- 614 structure shaped by hydroelectric power plants in the upper Rhine catchment. *Evolutionary*615 *Applications*, 9(2), 394-408. doi: 10.1111/eva.12339
- Hague, M. T. J., & Routman, E. J. (2016). Does population size affect genetic diversity? A test with
 sympatric lizard species. *Heredity*, *116*(1), 92-98. doi: 10.1038/hdy.2015.76
- Hand, B. K., Flint, C. G., Frissell, C. A., Muhlfeld, C. C., Devlin, S. P., Kennedy, B. P., ... Stanford, J. A.
- 619 (2018). A social-ecological perspective for riverscape management in the Columbia River
- Basin. Frontiers in Ecology and the Environment, 16(S1), S23-S33. doi: 10.1002/fee.1752
- Hawkins, P. R., Hortle, K. G., Phommanivong, S., & Singsua, Y. (2018). Underwater video monitoring
- 622 of fish passage in the Mekong River at Sadam Channel, Khone Falls, Laos. *River Research and*
- 623 Applications, 34(3), 232-243. doi: 10.1002/rra.3239
- Hedrick, P. W. (2005). A Standardized Genetic Differentiation Measure. *Evolution*, *59*(8), 1633-1638.
- 625 doi: 10.1111/j.0014-3820.2005.tb01814.x
- Jansson, R., Nilsson, C., & Malmqvist, B. (2007). Restoring freshwater ecosystems in riverine
- 627 landscapes : The roles of connectivity and recovery processes. *Freshwater Biology*, 52(4),

628 589-596. doi: 10.1111/j.1365-2427.2007.01737.x

- Januchowski-Hartley, S. R., Diebel, M., Doran, P. J., & McIntyre, P. B. (2014). Predicting road culvert
- 630 passability for migratory fishes. *Diversity and Distributions*, 20(12), 1414-1424. doi:
- 631 10.1111/ddi.12248
- Jombart, T., Devillard, S., & Balloux, F. (2010). Discriminant analysis of principal components : A new
- 633 method for the analysis of genetically structured populations. *BMC Genetics*, *11*(1), 94. doi:
- 634 10.1186/1471-2156-11-94

535 Jost, L. (2008). G_{ST} and its relatives do not measure differentiation. *Molecular Ecology*, 17(18),

636 4015-4026. doi: 10.1111/j.1365-294X.2008.03887.x

- 537 Junge, C., Museth, J., Hindar, K., Kraabøl, M., & Vøllestad, L. A. (2014). Assessing the consequences of
- habitat fragmentation for two migratory salmonid fishes. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 24(3), 297-311. doi: 10.1002/aqc.2391
- 640 Kimura, M. (1983). The Neutral Theory of Molecular Evolution. Cambridge University Press.
- 641 Klütsch, C. F. C., Maduna, S. N., Polikarpova, N., Forfang, K., Aspholm, P. E., Nyman, T., ... Hagen, S. B.
- 642 (2019). Genetic changes caused by restocking and hydroelectric dams in demographically
- 643 bottlenecked brown trout in a transnational subarctic riverine system. *Ecology and Evolution*,
- 644 ece3.5191. doi: 10.1002/ece3.5191
- Landguth, E. L., Cushman, S. A., Schwartz, M. K., McKELVEY, K. S., Murphy, M., & Luikart, G. (2010).
- Quantifying the lag time to detect barriers in landscape genetics. *Molecular Ecology*, *19*(19),
 4179-4191. doi: 10.1111/j.1365-294X.2010.04808.x
- 648 Li, Y.-C., Korol, A. B., Fahima, T., Beiles, A., & Nevo, E. (2002). Microsatellites : Genomic distribution,
- 649 putative functions and mutational mechanisms: a review. *Molecular Ecology*, 11(12),
- 650 2453-2465. doi: 10.1046/j.1365-294X.2002.01643.x
- Liaw, A., & Wiener, M. (2002). Classification and Regression by randomForest. *R News*, 2(3), 18-22.
- Lowe, W. H., & Allendorf, F. W. (2010). What can genetics tell us about population connectivity?

653 *Molecular Ecology*, *19*(15), 3038-3051. doi: 10.1111/j.1365-294X.2010.04688.x

- McLaughlin, R. L., Smyth, E. R. B., Castro-Santos, T., Jones, M. L., Koops, M. A., Pratt, T. C., & Vélez-
- Espino, L.-A. (2013). Unintended consequences and trade-offs of fish passage. *Fish and*
- 656 *Fisheries, 14*(4), 580-604. doi: 10.1111/faf.12003
- McNeish, D. M. (2014). Analyzing Clustered Data with OLS Regression : The Effect of a Hierarchical
 Data Structure. 40, 6.
- 659 McRae, B. H. (2006). Isolation by resistance. *Evolution*, 60(8), 1551–1561.

660 Meirmans, P. G. (2006). Using the Amova Framework to Estimate a Standardized Genetic

- 661 Differentiation Measure. *Evolution*, 60(11), 2399-2402. doi: 10.1111/j.0014-
- 662 3820.2006.tb01874.x
- Meirmans, P. G., & Hedrick, P. W. (2011). Assessing population structure : FST and related measures.
 Molecular Ecology Resources, *11*(1), 5-18. doi: 10.1111/j.1755-0998.2010.02927.x
- 665 Neuenschwander, S., Michaud, F., & Goudet, J. (2019). QuantiNemo 2 : A Swiss knife to simulate
- 666 complex demographic and genetic scenarios, forward and backward in time. *Bioinformatics*,
- 667 *35*(5), 886-888. doi: 10.1093/bioinformatics/bty737
- 668 Nilsson, C. (2005). Fragmentation and Flow Regulation of the World's Large River Systems. Science,
- 669 *308*(5720), 405-408. doi: 10.1126/science.1107887
- 670 Paz-Vinas, I., Comte, L., Chevalier, M., Dubut, V., Veyssiere, C., Grenouillet, G., ... Blanchet, S. (2013).
- 671 Combining genetic and demographic data for prioritizing conservation actions : Insights from 672 a threatened fish species. *Ecology and Evolution*, *3*(8), 2696-2710. doi: 10.1002/ece3.645
- 673 Paz-Vinas, I., Quéméré, E., Chikhi, L., Loot, G., & Blanchet, S. (2013). The demographic history of
- 674 populations experiencing asymmetric gene flow : Combining simulated and empirical data.
- 675 *Molecular Ecology*, 22(12), 3279-3291. doi: 10.1111/mec.12321
- 676 Poff, N. L., & Schmidt, J. C. (2016). How dams can go with the flow. Science (New York, N.Y.),
- 677 *353*(6304), 1099-1100. doi: 10.1126/science.aah4926
- Poulet, N. (2007). Impact of weirs on fish communities in a piedmont stream. *River Research and Applications*, 23(9), 1038-1047. doi: 10.1002/rra.1040
- 680 Pracheil, B. M., Mestl, G. E., & Pegg, M. A. (2015). Movement through Dams Facilitates Population
- 681 Connectivity in a Large River. *River Research and Applications*, *31*(5), 517-525. doi:
- 682 10.1002/rra.2751
- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using
 multilocus genotype data. *Genetics*, 155(2), 945-959. (WOS:000087475100039).

685	Prunier, J. G., Dubut, V., Chikhi, L., & Blanchet, S. (2017). Contribution of spatial heterogeneity in
686	effective population sizes to the variance in pairwise measures of genetic differentiation.
687	Methods in Ecology and Evolution, 8(12), 1866-1877. doi: 10.1111/2041-210X.12820

688 Prunier, J. G., Dubut, V., Loot, G., Tudesque, L., & Blanchet, S. (2018). The relative contribution of

- 689 river network structure and anthropogenic stressors to spatial patterns of genetic diversity in
- 690 two freshwater fishes : A multiple-stressors approach. *Freshwater Biology*, *63*(1), 6-21. doi:
- 691 10.1111/fwb.13034
- 692 R Development Core Team. (2014). *R: A Language and Environment for Statistical Computing, R*

693 Foundation for Statistical Computing. Consulté à l'adresse http://www.R-project.org

694 Raeymaekers, J. A. M., Raeymaekers, D., Koizumi, I., Geldof, S., & Volckaert, F. A. M. (2009).

- 695 Guidelines for restoring connectivity around water mills : A population genetic approach to
- the management of riverine fish. *Journal of Applied Ecology*, 46(3), 562-571. doi:
- 697 10.1111/j.1365-2664.2009.01652.x
- Reid, A. J., Carlson, A. K., Creed, I. F., Eliason, E. J., Gell, P. A., Johnson, P. T. J., ... Cooke, S. J. (2018).
- 699 Emerging threats and persistent conservation challenges for freshwater biodiversity.

700 Biological Reviews. doi: 10.1111/brv.12480

- Richardson, J. L., Brady, S. P., Wang, I. J., & Spear, S. F. (2016). Navigating the pitfalls and promise of
 landscape genetics. *Molecular ecology*, 25(4), 849–863.
- Rousset, F. (2008). GENEPOP '007 : A complete re-implementation of the GENEPOP software for
 Windows and Linux. *Molecular Ecology Resources*, 8(1), 103-106. (WOS:000253827100016).

705 Roy, S. G., Uchida, E., de Souza, S. P., Blachly, B., Fox, E., Gardner, K., ... Hart, D. (2018). A multiscale

- approach to balance trade-offs among dam infrastructure, river restoration, and cost.
- 707 Proceedings of the National Academy of Sciences, 115(47), 12069-12074. doi:
- 708 10.1073/pnas.1807437115
- Saint-Pé, K., Blanchet, S., Tissot, L., Poulet, N., Plasseraud, O., Loot, G., ... Prunier, J. G. (2018). Genetic
- admixture between captive-bred and wild individuals affects patterns of dispersal in a brown

711 trout (*Salmo trutta*) population. *Conservation Genetics*, *19*(5), 1269-1279. doi:

712 10.1007/s10592-018-1095-2

- Schlötterer, C. (2000). Evolutionary dynamics of microsatellite DNA. *Chromosoma*, *109*(6), 365-371.
 doi: 10.1007/s004120000089
- 715 Schwartz, M., Luikart, G., & Waples, R. (2007). Genetic monitoring as a promising tool for
- 716 conservation and management. *Trends in Ecology & Evolution, 22*(1), 25-33. doi:
- 717 10.1016/j.tree.2006.08.009
- 718 Selkoe, K. A., Scribner, K. T., & Galindo, H. M. (2015). Waterscape Genetics—Applications of
- 719 Landscape Genetics to Rivers, Lakes, and Seas. In N. Balkenhol, S. A. Cushman, A. T. Storfer, &
- 720 L. P. Waits (Éd.), Landscape Genetics (p. 220-246). doi: 10.1002/9781118525258.ch13
- Silva, A. T., Lucas, M. C., Castro-Santos, T., Katopodis, C., Baumgartner, L. J., Thiem, J. D., ... Cooke, S.
- J. (2018). The future of fish passage science, engineering, and practice. *Fish and Fisheries*,
 19(2), 340-362. doi: 10.1111/faf.12258
- Song, C., Omalley, A., Roy, S. G., Barber, B. L., Zydlewski, J., & Mo, W. (2019). Managing dams for
- 725 energy and fish tradeoffs : What does a win-win solution take? *Science of The Total*

726 *Environment, 669,* 833-843. doi: 10.1016/j.scitotenv.2019.03.042

Städele, V., & Vigilant, L. (2016). Strategies for determining kinship in wild populations using genetic
data. *Ecology and Evolution*, 6(17), 6107-6120. doi: 10.1002/ece3.2346

729 Storfer, A., Murphy, M. A., Spear, S. F., Holderegger, R., & Waits, L. P. (2010). Landscape genetics :

730 Where are we now? *Molecular Ecology*, *19*(17), 3496-3514. doi: 10.1111/j.1365-

- 731 294X.2010.04691.x
- Turgeon, K., Turpin, C., & Gregory-Eaves, I. (2019). Dams have varying impacts on fish communities
- 733 across latitudes : A quantitative synthesis. *Ecology Letters*, ele.13283. doi: 10.1111/ele.13283
- 734 Wang, J. L. (2005). Estimation of effective population sizes from data on genetic markers.
- 735 Philosophical Transactions of the Royal Society B-Biological Sciences, 360(1459), 1395-1409.
- 736 (WOS:000231317800005).

737 Weir, B. S., & Cockerham, C. C. (1984). Estimating F-Statistics for the analysis of population structure.

738 *Evolution*, *38*(6), 1358-1370. (WOS:A1984TY40400017).

- Wilson, G. A., & Rannala, B. (2003). Bayesian inference of recent migration rates using multilocus
 genotypes. *Genetics*, *163*(3), 1177-1191. (WOS:000182046900029).
- 741 Winter, D. J. (2012). MMOD : An R library for the calculation of population differentiation statistics.
- 742 *Molecular Ecology Resources*, *12*(6), 1158-1160. doi: 10.1111/j.1755-0998.2012.03174.x
- 743 Yue, G. H., David, L., & Orban, L. (2007). Mutation rate and pattern of microsatellites in common carp
- 744 (Cyprinus carpio L.). *Genetica*, *129*(3), 329-331. doi: 10.1007/s10709-006-0003-8
- 745
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Box

748								
749	Box 1 : Guidelines for the use and the interpretation of the C_{INDEX} .							
750	The C_{INDEX} allows an individual and standardized quantification of the impact of artificial barriers on							
751	riverscape functional connectivity from snapshot measures of genetic differentiation. Here, we provide							
752	a guideline for practitioners:							
753	• Species: any freshwater species whose local effective population sizes are lower than 1000 can be							
754		considered.						
755	•	Obstacle: any obstacle whose age corresponds to a minimum of 10-15 generations and a maximum						
756		of 600 generations for the studied species can be considered.						
757	•	Sampling: populations are sampled in the immediate upstream and downstream vicinity of the						
758		obstacle, with a minimum of 20-30 individuals per populations.						
759	•	Genetic data: Individual genotypes are based on a set of polymorphic microsatellite markers.						
760	•	Computation: The C_{INDEX} is computed in R thanks to a user-friendly script provided in Appendix						
761		S11 (see also Appendix S10 for a walkthrough).						
762	•	Interpretation for $C_{INDEX} < 10\%$: A C_{INDEX} value lower than 10% (or whose 95% confidence						
763		interval includes the 10% threshold) indicates no gene flow between populations (total barrier						
764		effect), whatever the age of the obstacle of the effective size of populations.						
765	•	Interpretation for C_{INDEX} > 90%: A C_{INDEX} value higher than 90% (or whose 95% confidence						
766		interval includes the 90% threshold) indicates full genetic connectivity (no barrier effect), whatever						
767		the age of the obstacle of the effective size of populations.						
768	•	Interpretation for intermediate C_{INDEX} values: Intermediate C_{INDEX} values can be used to rank						
769		obstacles according to their barrier effect. However, for C_{INDEX} values ranging from ~15 to ~70%,						
770		the C_{INDEX} tends to slightly increase with both the increase in time since barrier creation and the						
771		increase in effective population sizes (Figure 3I-J).						
772	•	Obstacles with C_{INDEX} values that do not differ by more than 15 to 20% but that are characterized						
773		by very different ages and / or population sizes (as indicated by expected heterozygosity) should be						

considered as possibly having comparable barrier effects, except of course if the ranking of obstacles based on C_{INDEX} values goes against these trends. Consider for instance an obstacle A of age 20 (in generations) and an obstacle B of age 300. If $C_{INDEX}(A) = 20\%$ and $C_{INDEX}(B) = 40\%$, both obstacles should be considered as possibly having the same impact on gene flow. On the contrary, if $C_{INDEX}(A) = 40\%$ and $C_{INDEX}(B) = 20\%$, obstacle B can be confidently considered as more impactful than obstacle A.

781 **Table**

Table 1. Main characteristics and results for the obstacles selected from empirical datasets (Original publication: (1) Gouskov et al., 2016; (2) Prunier et al., 2018). For each obstacle, the table indicates the name of the river, the date of creation, the distance between upstream and downstream sampled populations, the considered species (*Sc: Squalius cephalus*; *Go* : *Gobio occitaniae*; *Pp: Phoxinus phoxinus*), the number of generations elapsed since barrier creation, the mean expected heterozygosity (*He*), and the computed C_{INDEX} along with its 95% confidence inverval. In bold, obstacles that were found as significant barriers to gene flow in the considered species.

River	Obstacle	Creation date	Upstream- Downstream distance (km)	Species	Number of elapsed generations	Не	C _{INDEX}	95%CI	Original publication
Rhine	Barr05	1966	12.02	Sc	14.67	0.76	85.50	16.53	(1)
Rhine	Barr06	1914	8.28	Sc	32.00	0.75	100	0	(1)
Rhine	Barr07	1933	7.19	Sc	25.67	0.75	100	0	(1)
Rhine	Barr08	1941	12.86	Sc	23.00	0.73	88.51	2.05	(1)
Rhine	Barr09	1920	6.38	Sc	30.00	0.73	90.02	4.31	(1)
Rhine	Barr10	1956	12.42	Sc	18.00	0.69	100	0	(1)
Rhine	Barr11	1964	4.79	Sc	15.33	0.69	74.98	13.82	(1)
Aar	Barr13	1902	1.91	Sc	36.00	0.76	64.21	8.21	(1)
Aar	Barr14	1953	10.75	Sc	19.00	0.76	63.59	5.58	(1)
Aar	Barr15	1945	7.50	Sc	21.67	0.75	86.49	15.39	(1)
Aar	Barr16	1929	5.61	Sc	27.00	0.77	100	0	(1)
Aar	Barr17	1893	3.14	Sc	39.00	0.77	100	0	(1)
Aar	Barr18	1917	15.55	Sc	31.00	0.76	96.60	4.09	(1)
Aar	Barr19	1896	1.93	Sc	38.00	0.76	74.49	4.06	(1)
Aar	Barr20	1896	6.82	Sc	38.00	0.75	69.41	3.90	(1)
Aar	Barr21	1970	6.00	Sc	13.33	0.76	100	0	(1)
Aar	Barr22	1970	9.51	Sc	13.33	0.77	100	0	(1)
Aar	Barr23	1939	9.22	Sc	23.67	0.76	100	0	(1)
Aar	Barr24	1900	11.67	Sc	36.67	0.76	100	0	(1)
Aar	Barr25	1968	5.81	Sc	14.00	0.76	100	0	(1)
Aar	Barr26	1963	4.84	Sc	15.67	0.75	100	0	(1)
Limmat	Barr32	1933	16.08	Sc	25.67	0.73	71.30	2.10	(1)
Limmat	Barr33	1933	3.24	Sc	25.67	0.72	100	0	(1)
Célé	CLA	1500	0.18	Go	204	0.60	82.03	5.28	(2)
Célé	SCA	1500	0.09	Go	204	0.63	92.73	3.88	(2)
Célé	SCC	1960	0.2	Go	20	0.64	49.23	3.78	(2)
Viaur	SEG	1600	0.11	Go	164	0.58	100	0	(2)
Viaur	CAM	1600	0.49	Go	164	0.62	100	0	(2)
Viaur	CAP	1700	0.55	Go	124	0.61	83.85	1.33	(2)
Viaur	SJU	1800	1.07	Go	64	0.62	81.98	3.98	(2)
Viaur	CIR	1960	0.97	Go	20	0.62	86.72	15.10	(2)
Célé	CLA	1500	0.18	Pp	255	0.54	90.29	5.14	(2)
Célé	SCA	1500	0.09	Pp	255	0.57	100	0	(2)
Célé	SCC	1960	0.2	Pp	25	0.58	100	0	(2)
Viaur	SEG	1600	0.11	Pp	205	0.63	100	0	(2)
Viaur	CAM	1600	0.49	Pp	205	0.61	100	0	(2)
Viaur	CAP	1700	0.55	Pp	155	0.67	100	0	(2)
Viaur	SJU	1800	1.07	Pp	105	0.70	100	0	(2)
Viaur	CIR	1960	0.97	Pp	25	0.70	100	0	(2)
Célé	CLA	1500	0.18	Go-Pp	/	/	86.16	8.10	(2)
Célé	SCA	1500	0.09	Go-Pp	/	/	96.37	7.12	(2)
Célé	SCC	1960	0.2	Go-Pp	/	/	74.61	49.76	(2)
Viaur	SEG	1600	0.11	Go-Pp	/	/	100	0	(2)
Viaur	CAM	1600	0.49	Go-Pp	/	/	100	0	(2)
Viaur	CAP	1700	0.55	Go-Pp	/	/	91.93	15.83	(2)
Viaur	SJU	1800	1.07	Go-Pp	/	/	90.99	17.66	(2)
v iaur	CIK	1960	0.97	GO-PD	/	/	93.30	15.02	(2)

Figures 794





797 Figure 1. Flowchart illustrating the major steps in calculating the index of genetic connectivity for two 798 independent obstacles. This flowchart refers to a user-friendly script provided in Appendix S11 (see 799 also Appendix S10 for a walkthrough). After the sampling of populations located at the immediate 800 upstream and downstream vicinity of each obstacle, users only have to provide a file of genotypes in the genepop format and a file of parameters indicating, for each obstacle, the names of the sampled 801 802 populations and the number T of generations elapsed since the creation of the obstacle. Observed 803 measures of genetic differentiation GD_{obs} and mean expected heterozygosity He are automatically computed from provided genotypic data. GD_{min} and GD_{max} values, both delimiting the theoretical 804 range of variation of GD_{obs} , are automatically predicted from pre-existing .rda files, GD_{max} values 805 depending on both He and T. The computation of the index basically amounts to rescaling GD_{obs} 806

within its theoretical range (see main text for details), thus allowing standardized comparisons of the
permeability of various obstacles, whatever their age, the considered species or the effective size of
sampled populations.



Figure 2. For each mutation rate (panels A and B) and each metric of genetic differentiation (G''st on the left and φ 'st on the right), predicted GD_{max} variations across the parameter space defined by the time *T* elapsed since total barrier creation (from 0 to 600 generations) and the averaged expected heterozygosity (*He*, a proxy for effective population size, ranging from 0 to 0.93) for pairs of adjacent populations. GD_{min} values are represented at the bottom of each graph. GD_{min} and GD_{max} surfaces together delimit the theoretical range of variation for any observed measure of genetic differentiation GD_{obs} .





822 Figure 3. Panels A-H: C_{INDEX} responses to the increase in migration rate (m, on a logarithmic scale) for 823 five different population sizes Ne (colored lines) and from 5 to 300 generations after barrier creation 824 (panels A to H). All C_{INDEX} values were averaged over 20 simulated replicates and plotted with 95% 825 confidence intervals. In each panel, the shaded pink surface and the black arrow indicate the range of 826 variation of C_{INDEX} values for a migration rate of 0.05 (mean value across effective population sizes, 827 minimum and maximum values into brackets). Panel I-J: CINDEX responses to the increase in time since 828 barrier creation (panel I) and to the increase in effective population size (panel J) for eight different 829 migration rates m (colored lines). The mean C_{INDEX} values computed over simulated replicates were 830 here averaged over effective population sizes (panel I) or over generations (excluding generations \leq 831 10; panel J) and plotted with standard deviations. In all panels, shaded grey areas represent the ranges of variations in which the monitored obstacle can be considered as acting as a total barrier to gene 832 833 flow ($C_{INDEX} < 10\%$) or, on the contrary, as allowing full genetic connectivity ($C_{INDEX} > 90\%$).



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Figure 4. *C*_{INDEX} values as computed from empirical datasets. Panel A: Results for the 23 dams selected from Gouskov et al. (2016), ranked according to their inferred impact on chubs' genetic connectivity (from the most impactful on the left to the less problematic ones on the right). Panel B: For each monitored species (Go: *Gobio occitaniae*; Pp: *Phoxinus phoxinus*), results for the 8 weirs selected from Prunier et al. (2018), ranked according to their date of creation (indicated by the number of generations elapsed since barrier creation in each species).

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